Yeastern Biotech Co., Ltd

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Ver. S0124

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Email: service@yb-biotech.com

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2X O'in1 DNA Polymerase Premix II w/blue dye

Cat. No.
FYT208-200P
SYT208-001

2X O'in1 DNA Polymerase Premix II w/blue dye

Specification: 200 preps,1ml x 2 **Storage:** -20°C

Description

2X O'in1 DNA Polymerase Premix II w/blue dye is an economical and ready-to-use premix, containing a high-sensitivity and high-yield YEAtaq DNA Polymerase, dNTP and all other reagents necessary for PCR, except DNA template and primers. It saves the time for preparing the master mix and reduces the risk of contamination from multiple pipetting steps. 2X O'in1 DNA Polymerase Premix II w/blue dye is also available with non-interfering dye for applications when loading dye is desired.

Optimal PCR conditions, including template, primer concentrations and PCR program, for gene of interest should be determined experimentally by the investigator from case to case.

Application

1. PCR products for TA Cloning
2. Trace amount of DNA detection
3. Suitable for economic screening
4. High throughput PCR
5. Rutine PCR with high reproducibility
6. Amplify target gene from genomic DNA

Size of Dye (in 1% agarose gel)

Blue dye : ~500 bp

Unit Definition

One unit is defined as the amount of enzyme that will incorporate 10 nmol of dNTPs into acid-insoluble material in 30 minutes at 72°C.

Procedure

A. Preparation of the PCR Mixture

1. Prepare a master mixture according to the Table below.

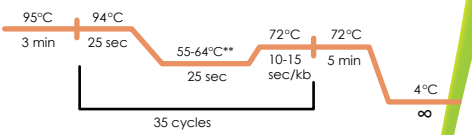
Component	Volume	Final conc.
2× O'in1 DNA Polymerase Premix II (blue)	10 µl	1×
Forward Primer (10 µM)	0.5 µl	0.25 µM*
Reverse Primer (10 µM)	0.5 µl	0.25 µM*
Template DNA	0.5-5 µl	<1 µg
ddH_2O	total to 20 µl	

*The final concentration of the primers can be adjusted according to the experiment.

2. Mix the master mixture thoroughly by pipetting up and down. Dispense the mixture into PCR tubes or plates.

B. Performing PCR

1. Program your instrument according to the graph below.



**The optimal annealing temperature can be adjusted according to primer's Tm value.

2. Place the PCR tubes or PCR plates in the thermal cycler and start the cycling program.

3. Load samples on agarose gel and perform electrophoresis.

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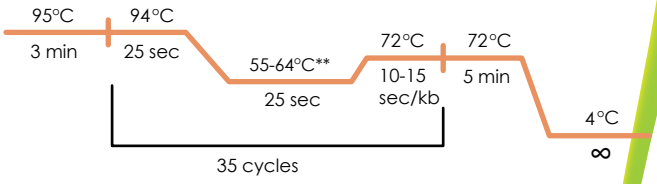

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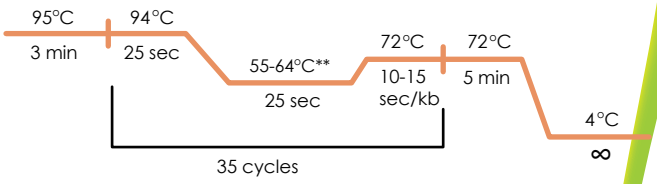

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